Autonomous Micron-scale system for *in vivo* cell monitoring



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Trends in microsystems for *in vivo* monitoring



Node	Maturity	Power (W)	Size(m ³)
Obtrusive	Commercial	1-10 ⁻¹	10 ⁻³
Parasitic	Prototype/Comm.	10 ⁻² -10 ⁻³	10 ⁻⁶
Symbiotic	Research/Protot.	10 ⁻⁵ – 10 ⁻⁶	10 ⁻⁹
Bio-hybrid	Concept/Research	< 10 ⁻⁷	10 ⁻¹⁵

Molecular targets for *in vivo* monitoring of the cell

Extract **mRNA** to monitor **genetic expression** is the most powerful way to be able to address complex questions on the cell state.

Micron-Scale System for *in vivo* genetic expression analysis and drug delivery

- symbiotic to bio-hybrid node
 multi-site
- heterogeneous integration

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Components of an autonomous microsystem for *in vivo* cell expression

Heterogeneous Integration

Bio-interface

(mRNA sample extraction, handling, modification)

Sensing/Transduction

(Sensing physical-chemical elements, measurement)

Actuation (Drug delivery)

Communication/Power Processing and storage

(Energy harvester, Communication module) (Data elaboration for monitoring or actuation)

Bio-interface. mRNA extraction

mRNA extraction from cells

Cell isolation



Cell lysis





J.W. Hong *et al., Nature Biotechnology, (2004) 22-4, pp. 435-439.*

- **Device:** PDMS by soft lithography.
- Sample processing: Pressure and reagents
- Biological sample: Measurable RNA was extracted from down to 28 cells processed in a 0.4 nl volume.

• Time for processing: several minutes

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Bio-interface. mRNA extraction

Scaling trade-off

mRNA extraction and analysis from SINGLE CELL?

- Technically feasible, but
- Single cell ⇒ Single molecules. The minimum quantity of final mRNA depends on the sensor performance.

This limit can be relaxed by the integration of an Amplification Reactor

Bio-interface. Amplification

Polymerase Chain Reaction

Thermal cycling between Ts. A, B, C (95°C, 77°C, 60°C).

A - 95°C Separation of original molecules

B – 77° C Strands binds to suitable primers for targeted amplification.

C - The polymerase enzyme extends "builds" double strands from single elements C. Guiducci SRC Forum – Stanford University



M.U. Koop *et al., Science (1998), 280, pp1046-1047*

• Device: Glass and copper

 Sample processing: reagents (enzymes, primers, oligonucleotides)

 Biological sample: few nl, 2²⁰ amplification factor.

• Time for processing: 18 min

Bio-interface. Amplification

Heat management

• Low thermal mass of the sample and high surface-to-volume ratio allows for rapid temperature cycling between temperature setpoints.

- Heating and cooling time are each less than 100ms (90 μ m × 40 μ m channels in glass and copper heaters).

Thermal coatings may impact cooling schedule.

Energy costs Costs of thermal cycling on chip

 Power consumption for heating can be estimated to be 1-10mW for a flow rate 100 nl/sec. (D.J. Saldler, *IEEE Trans. On Components* and Pack. Tech. (2003), 26-2, pp. 309-316)
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Sensing/Transduction. Techniques



Sensing/Transduction. Electrical Technique based on capacitance measurements

128-site CMOS chip

 Capacitance measurement by capacitance to frequency conversion.

Fully digital output

 Reduced number of pads by on chip addressing



C. Stagni *et al., IEEE* J. of Solid-State Circuits (2006) 41-12, pp. 2956-2964

250µm

- Device: Gold on CMOS standard process
- Sample processing: hybridization reaction at RT
- Time for processing: 15 min (biochemical reaction)

Biological sample: 500 nl of 10nM (10⁷
 C. Guiducci SRC Forum – Stanford University molecules per site)
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Sensing/Transduction. Electrical Technique based on capacitance measurements

Scaling Trade-off *Minimum dimension of the spot* • Capacitance signal for a 200 μ m spot and 10² molecules/ μ m²: 0.9 nF ± 0.01 nF (30 fF/ μ m²). • Limit of the spot size for 6dB SNR capacitance signal \rightarrow 15 μ m

Energy costs

Cost of measurement per site

• Input generation: $10 \text{ mV} \cdot 1 \text{ mA} = 10 \mu \text{W}$

Sample rate: 1Hz, 1 sec per site

0.01 nF

 Measurement: 100 µW (for low power amplifiers). Need for on-chip multiplexing strategies.

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15µm

Processing. Electrical Technique based on capacitance measurements

Energy costs Cost of the data processing for sensing output

 1000 instructions per site to perform averages and thresholding.

1MIPS microprocessor. Power consumption range from ultra low power to standard:
 [2.6 pJ/instr – 20 pJ/instr] →
 [2.6 nW per site – 20 nW per site].

• In case of 1000 site the power consumption ranges from [2.6 μW – 20 μW].

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Communication

NO CON

Communication

Wireless data communication for monitoring purposes

Energy cost per site

- 12 bit/site per second (1 site per second)
- down to 10⁻⁷J/bit with RF systems (J. Ammer, *IWWAN'06*)
- \Rightarrow 1 µW per site
- In case of 1000 site the power consumption for communication is 1 mW.

Actuation

Local delivery of controlled quantity of drugs

 Array of reservoirs in an electrolyte impermeable substrate

 Gold membranes (300nm) that close the reservoirs are anodes and other gold electrodes are cathodes.
 Surface of membranes: 50
 × 50 µm²

 Membranes dissolve at specific potential applied
 1.04 V and 3 µA. P = 3 µ W



J.T. Santini, Angewandte (2000), 39, pp. 2396-2407

- Device: Bulk micromachining of Si process.
- Sample processing: drug release one well at a time.
- Time for processing: 30 sec per well
- Biological sample: 25 nl

Total Energy costs for a node with monitoring purposes.

- Bio-interface: 10mW, determined by PCR amplification reaction.
- Power cost per site for signal generation and

communication

110 μW. Sensing and transduction, determined by amplifier consumption.

- 10 nW. Processing.
- 1 µW. Communication

Power cost for 1000 sites for signal generation and communication

110 mW without multiplexing strategies.

Bottle neck: PCR and analog measurement circuit without multiplexing strategies.

Total Energy costs for a node for monitoring and actuation

Bio-interface (non PCR-free)

	Bio- interface	Signal Transduction	Processing	Communication	Actuation
Per site	1-10mW	110µW	10nW	1µW	10µW
100 sites	1-10mW	11mW	1µW	100µW	10µW
1000 sites	1-10mW	110mW	10µW	1mW	10µW

Processing for control generation non critical

Wireless Power Delivery

- Standard UHF RFID energy trasfer is about 40 µW
- More advanced implementation can reach up to 1mW (Powercast) at 1 m distance.
 - Suitable for PCR free, serial-mode or low parallelism analysis devices

- Inductive power coupling up to hundreds of mW.
 - Suitable for PCR-based, parallel-mode systems